

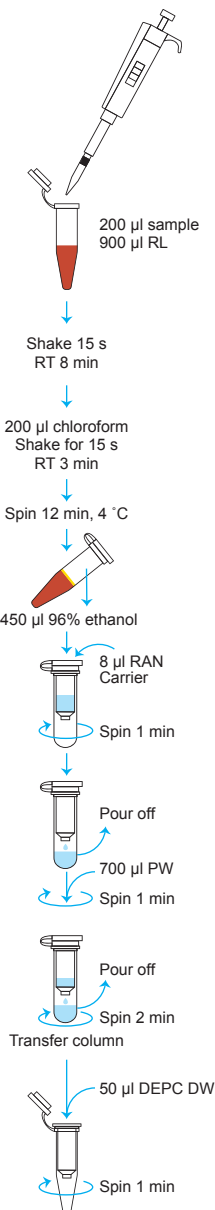
## Before Starting

Add 48 ml of absolute ethanol to the PW (only at the first use).

## Reagents NOT Provided

1. Chloroform
2. 96% ethanol

## Protocol



- 1 Transfer 200  $\mu$ l of sample whole blood or buffy coat (to get more RNA) to a 1.5 ml tube and add 900  $\mu$ l of RL solution. Shake it for 15 sec and incubate at room temperature for 8 min. Shake it each 4 min to help lysis process.
- 2 Add 200  $\mu$ l of chloroform to the mixture. Shake it completely for 15 s and incubate for 3 min at room temperature.
- 3 Spin for 12 min at 13,000 rpm at 4 °C.
- 4 Transfer 450  $\mu$ l of the upper phase into a new 1.5 ml tube. Add 8  $\mu$ l RNA carrier and mix it completely for 5 sec. Add 450  $\mu$ l of 96% ethanol to the mixture and mix them well.
- 5 Transfer mixture to the spin column. Do NOT touch upper rim of column. Spin for 1 min at 13,000 rpm.
- 6 Pour off the flow-through of collection tube.
- 7 Add 700  $\mu$ l of PW and spin for 1 min at 13,000 rpm.
- 8 Pour off the flow-through of collection tube. (Optional: repeat step 8 and 9 with 500  $\mu$ l of PW to have more pure RNA)
- 9 Spin for 2 min at 13,000 rpm to remove the remaining of the wash buffer. Transfer the spin column to a new 1.5 ml microtube.
- 10 Add 50  $\mu$ l of DEPC-treated water, wait 3 min at room temperature or 57 °C (For more yield). If you want more concentration add less DEPC-treated water (35  $\mu$ l).
- 11 Spin for 1 min at 13,000 rpm to elute RNA from the column. Store RNA solution at -70 °C.

## Blood RNA Extraction Kit

### Before Starting

Add 48 ml of absolute ethanol to the PW (only at the first use).

### Reagents NOT Provided

1. Chloroform
2. 96% ethanol

### kit content

No.	Description:	Packing	Qty.
1	RL Buffer (RNA Lysis Buffer)	45 ml	1
2	PW Buffer (Wash Buffer)	12 ml	1
3	DEPC-treated Water	3 ml	1
4	RNA Carrier	300 ul	1
5	Spin Column	50	1
6	Collection Tube	50	1